

## Two New Abietane Diterpenoids from the Stems of *Clerodendrum kaichianum* P. S. HSU

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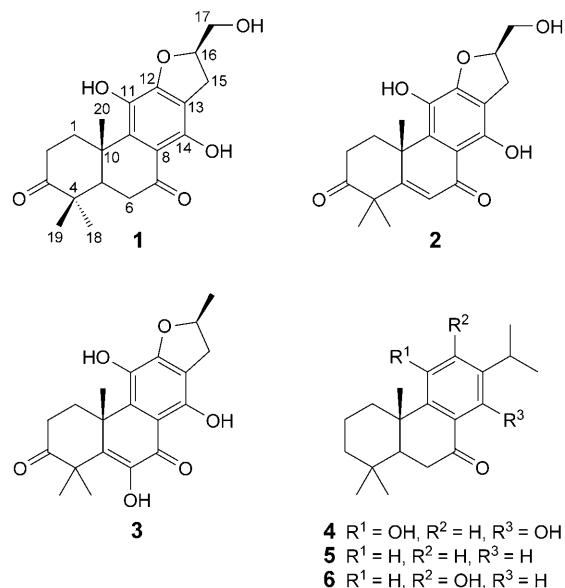
Two new abietane diterpenoids, named 17-hydroxyteuvinenone G (**1**) and 17-hydroxyteuvinenone-5(6)-enone G (**2**), together with four known diterpenoids, were isolated from the stems of *Clerodendrum kaichianum* P. S. Hsu. Their structures were elucidated by extensive NMR and MS analyses, and by comparison with literature data. The new compounds showed significant cytotoxicities against the HL-60 and A-549 tumor cell lines.

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**Introduction.** – The genus *Clerodendrum* contains more than 30 species distributed in China, some of which have been used in Traditional Chinese Medicine, such as *Clerodendrum indicum* for treating malaria and rheumatism [1]. Plants of the genus *Clerodendrum* have proved to be a rich source of abietane diterpenoids, as well as iridoids, triterpenes, phenylethanoid glycosides, and saponins, which have been found to possess many beneficial pharmacological effects, such as antimalarial, antitumor, and anti-HIV activities [2–8]. In China, the leaves of *Clerodendrum kaichianum* P. S. Hsu are used as a traditional medicine against hypertension. Search for the new bioactive natural products from this plant has led to the isolation of two new abietane diterpenoids, **1** and **2**, together with four known abietane diterpenoids, teuvinenone A (**3**), 11,14-dihydroxyabieta-8,11,13-trien-7-one (**4**), dehydroabietan-7-one (**5**), and sugiol (**6**), and these two new compounds were found to show cytotoxicity against HL-60 and A-549 cells. Here, we describe the isolation, structure elucidation, and cytotoxic activity of the new compounds.

**Results and Discussion.** – The 75% aqueous EtOH extract of the air-dried stems of *C. kaichianum* was suspended in H<sub>2</sub>O, and then partitioned with petroleum ether (PE), AcOEt, and BuOH successively. The AcOEt-soluble fraction was further chromatographed on silica gel and *Sephadex LH-20*, and preparative HPLC afforded diterpenoids **1–6**.

17-Hydroxyteuvinenone G (**1**) was obtained as yellowish needles and assigned the molecular formula C<sub>20</sub>H<sub>24</sub>O<sub>6</sub> on the basis of HR-ESI-MS ( $m/z$  359.1499 ( $[M - H]^-$ , C<sub>20</sub>H<sub>23</sub>O<sub>6</sub>; calc. 359.1493), which indicated nine degrees of unsaturation. The IR spectrum showed the absorption bands for OH (3420 cm<sup>-1</sup>), ketones (1714, 1678 cm<sup>-1</sup>), and benzene moieties (3034, 1612, 1580, 1508 cm<sup>-1</sup>). The absorption bands in the UV spectrum (235, 295, and 380 nm) also indicated the presence of a benzene ring and a ketone.



The <sup>1</sup>H-NMR spectrum (*Table 1*) of **1** showed signals corresponding to three Me groups ( $\delta(\text{H})$  1.17, 1.16, and 1.42 (*s*, 3 H each)) at quaternary C-atoms, which were almost identical to those of teuvinconone G assigned by *Cuadrado et al.* [9] to the Me(18), Me(19) and Me(20) groups ( $\delta(\text{H})$  1.18, 1.17 and 1.44, resp.). Signal corresponding to three OH groups (*singlets* at  $\delta(\text{H})$  13.15, 6.30, and 5.39), the strongly deshielded resonance of an OH H-atom ( $\delta(\text{H})$  13.15), together with its slow exchange with D<sub>2</sub>O, confirmed the existence of a phenolic OH group at C(14), which forms a strong intramolecular H-bond with the C(7)=O function.

In total, 20 C-atom signals were observed in the <sup>13</sup>C-NMR and DEPT 135 spectra of **1** (*Table 1*), including two C=O signals at  $\delta(\text{C})$  216.0 and 203.1, and six aromatic C-atom signals at  $\delta(\text{C})$  155.5, 155.3, 137.3, 131.4, 111.1, and 110.2. The high-field region displayed three Me signals at  $\delta(\text{C})$  17.5, 20.9, and 27.0, five CH<sub>2</sub> signals at  $\delta(\text{C})$  28.7, 34.4, 35.3, 35.7, and 64.7, two CH signals at  $\delta(\text{C})$  49.1 and 86.6, and two signals at ( $\delta(\text{C})$  39.3 and 46.9) for quaternary C-atoms (*Table 1*). These data revealed that compound **1** was a diterpenoid. The NMR spectra of **1**, being very similar to those of teuvinconone G [8], which showed the presence of a dihydro-2-(hydroxymethyl)furan fused with the aromatic ring ( $\delta(\text{H})$  3.35 (*dd*,  $J=15.0, 9.0$ , H <sub>$\alpha$</sub> -C(15)), 3.31 (*dd*,  $J=15.3, 7.0$ , H <sub>$\beta$</sub> -C(15)); 5.13–5.17 (*m*, H-C(16)); 3.90 and 3.80 (*dd*, H <sub>$\alpha$</sub> -C(17) and H <sub>$\beta$</sub> -C(17));  $\delta(\text{C})$  28.7 (*t*, C(15)); 86.6 (*d*, C(16)); 64.7 (*t*, C(17))) instead of the dihydro-2-methylfuran moiety in teuvinconone G ( $\delta(\text{H})$  3.38 and 2.85 (*dd*,  $J_{\text{gem}}=15.3$ , H <sub>$\alpha$</sub> -C(15) and H <sub>$\beta$</sub> -C(15)); 5.14 (*m*, H-C(16)); 1.52 (*d*, Me(17));  $\delta(\text{C})$  34.3 (*t*, C(15)); 83.5 (*d*, C(16)); 22.0 (*q*, C(17))).

Based on the analyses of the <sup>1</sup>H,<sup>1</sup>H-COSY and HSQC data, a *ABX* system ( $\delta(\text{H})$  2.41 (*dd*); 2.60–2.63 (*m*); 2.60–2.63 (*m*)) due to the position 5 and 6 of ring *B* was deduced, indicating the presence of the following fragments: CH<sub>2</sub>(1)–CH<sub>2</sub>(2),

Table 1.  $^{13}\text{C}$ - (125 MHz) and  $^1\text{H}$ -NMR (500 MHz) Data<sup>a</sup> of Compound **1**. Recorded in  $\text{CDCl}_3$ ,  $\delta$  in ppm,  $J$  in Hz.

	$\delta(\text{C})$	$\delta(\text{H})$	HMBC (H $\rightarrow$ C)
$\text{H}_\alpha\text{-C}(1)$	35.7 ( <i>t</i> )	1.99 ( <i>ddd</i> , $J = 13.7, 9.7, 9.7$ )	C(2), C(3), C(5), C(20)
$\text{H}_\beta\text{-C}(1)$		3.28 ( <i>dd</i> , $J = 13.7, 7.0$ )	
$\text{H}_\alpha\text{-C}(2)$	34.4 ( <i>t</i> )	2.73 ( <i>dd</i> , $J = 14.5, 7.9$ )	C(1), C(3), C(4)
$\text{H}_\beta\text{-C}(2)$		2.60–2.63 ( <i>m</i> )	
C(3)	216.0 ( <i>s</i> )		
C(4)	46.9 ( <i>s</i> )		
H–C(5)	49.1 ( <i>d</i> )	2.41 ( <i>dd</i> , $J = 11.2, 7.5$ )	C(1), C(4), C(6), C(7), C(10)
$\text{H}_\alpha\text{-C}(6)$	35.3 ( <i>t</i> )	2.60–2.63 ( <i>m</i> )	C(5), C(7), C(10)
$\text{H}_\beta\text{-C}(6)$		2.60–2.63 ( <i>m</i> )	
C(7)	203.1 ( <i>s</i> )		
C(8)	110.2 ( <i>s</i> )		
C(9)	137.3 ( <i>s</i> )		
C(10)	39.3 ( <i>s</i> )		
C(11)	131.4 ( <i>s</i> )		
C(12)	155.3 ( <i>s</i> )		
C(13)	111.1 ( <i>s</i> )		
C(14)	155.5 ( <i>s</i> )		
$\text{H}_\alpha\text{-C}(15)$	28.7 ( <i>t</i> )	3.35 ( <i>dd</i> , $J = 15.3, 9.0$ )	C(12), C(13), C(14), C(16)
$\text{H}_\beta\text{-C}(15)$		3.31 ( <i>dd</i> , $J = 15.3, 7.0$ )	
H–C(16)	86.6 ( <i>d</i> )	5.13–5.17 ( <i>m</i> )	C(13), C(15), C(17)
$\text{H}_\alpha\text{-C}(17)$	64.7 ( <i>t</i> )	3.90 ( <i>dd</i> , $J = 15.0, 8.7$ )	C(15), C(16)
$\text{H}_\beta\text{-C}(17)$		3.80 ( <i>dd</i> , $J = 15.0, 7.2$ )	
Me(18)	27.0 ( <i>q</i> )	1.17 ( <i>s</i> )	C(3), C(4), C(5), C(19)
Me(19)	20.9 ( <i>q</i> )	1.16 ( <i>s</i> )	C(3), C(5), C(18)
Me(20)	17.5 ( <i>q</i> )	1.42 ( <i>s</i> )	C(1), C(5), C(9), C(10)
11-OH		5.39 ( <i>br. s</i> )	
14-OH		13.15 ( <i>br. s</i> )	

<sup>a</sup>) Assignments were accomplished by a combination of 1D- and 2D- ( $^1\text{H}, ^{13}\text{C}$ -COSY, HSQC, and HMBC) NMR experiments.

$\text{CH}(5)\text{-CH}_2(6)$ , and  $\text{CH}_2(15)\text{-CH}(16)\text{-CH}_2(17)$  should exist. In the HMBC spectrum, the  $^1\text{H}, ^{13}\text{C}$  long-range correlations H–C(1)/C(2), C(3), C(5), C(10), and C(20); H–C(5)/C(1), C(3), C(4), C(6), C(7), and C(10); H–C(17)/C(15), and C(16); and H–C(15)/C(13), C(14), C(15), C(16), and C(17) (Table 1) suggested the abietane diterpenoid framework in the molecule. Furthermore, the  $^1\text{H}, ^{13}\text{C}$  long-range correlations H–C(18)/C(3), C(4), C(5), and C(19); H–C(19)/C(3), C(4), C(5), and C(18); and H–C(20)/C(1), C(5), C(9), and C(10) indicated that C(18) and C(19) were at C(4), and C(20) was at C(10). Therefore, compound **1** possesses an abietane-type diterpenoid framework with a  $\text{CH}_2\text{OH}$  group at C(16); the absolute configuration at C(16) was not ascertained. However, on biogenetic basis, it is reasonable to assume that it possess the same absolute configuration as the known abietane diterpenoid **3**, which was isolated from the same plant. Thus, the structure of **1** was suggested to be (16*R*)-12,16-epoxy-11,14,17-trihydroxy-17(15  $\rightarrow$  16)-*abeo*-abieta-8,11,13-triene-2,7-dione.

17-Hydroxyteuvinone **2** was obtained as yellowish needles and assigned the molecular formula  $C_{20}H_{22}O_6$  on the basis of HR-ESI-MS ( $m/z$  357.1330 ( $[M - H]^-$ ,  $C_{20}H_{21}O_6^-$ ; calc. 357.1336), which indicated ten degrees of unsaturation. The IR spectrum showed the absorption bands for OH ( $3402\text{ cm}^{-1}$ ) and ketone ( $1719$  and  $1672\text{ cm}^{-1}$ ) groups, and benzene moieties ( $1612$ ,  $1582$ , and  $1510\text{ cm}^{-1}$ ). The absorption bands in the UV spectrum ( $265$ ,  $296$ ,  $336$ , and  $382\text{ nm}$ ) also indicated the presence of a benzene ring.

The  $^{13}\text{C}$ -NMR and DEPT-NMR spectra indicated that **2** contained three Me, four  $\text{CH}_2$ , two CH groups, and eleven quaternary C-atoms. The NMR spectra of **2** were very similar to those of compound **1** except that the signals at  $\delta(\text{C})$  49.1 and 35.3 were missing, which were replaced by two signals at  $\delta(\text{C})$  174.1 and 122.4, indicating an additional  $\text{C}(5)=\text{C}(6)$  bond (Table 2). Thus, the structure of **2** was suggested to be (16*R*)-12,16-epoxy-11,14,17-trihydroxy-17(15  $\rightarrow$  16)-abeo-abieta-5,8,11,13-tetraene-2,7-dione.

Table 2.  $^{13}\text{C}$ - (125 MHz) and  $^1\text{H}$ -NMR (500 MHz) Data<sup>a</sup> of Compound **2**. Recorded in  $\text{CDCl}_3$ ,  $\delta$  in ppm,  $J$  in Hz.

	$\delta(\text{C})$	$\delta(\text{H})$	HMBC (H $\rightarrow$ C)
$\text{H}_\alpha\text{-C}(1)$	26.6 ( <i>t</i> )	1.96–2.01 ( <i>m</i> )	C(2), C(3), C(5), C(20)
$\text{H}_\beta\text{-C}(1)$		3.26 ( <i>dd</i> , $J = 14.5, 7.0$ )	
$\text{H}_\alpha\text{-C}(2)$	33.2 ( <i>t</i> )	2.70–2.73 ( <i>m</i> )	C(1), C(3), C(4)
$\text{H}_\beta\text{-C}(2)$		2.70–2.73 ( <i>m</i> )	
C(3)	213.2 ( <i>s</i> )		
C(4)	49.5 ( <i>s</i> )		
C(5)	174.2 ( <i>s</i> )		
H–C(6)	122.4 ( <i>d</i> )	6.30 ( <i>s</i> )	C(4), C(5), C(8), C(10)
C(7)	189.5 ( <i>s</i> )		
C(8)	108.9 ( <i>s</i> )		
C(9)	135.2 ( <i>s</i> )		
C(10)	41.6 ( <i>s</i> )		
C(11)	131.1 ( <i>s</i> )		
C(12)	154.0 ( <i>s</i> )		
C(13)	111.7 ( <i>s</i> )		
C(14)	154.1 ( <i>s</i> )		
$\text{H}_\alpha\text{-C}(15)$	28.8 ( <i>t</i> )	3.31 ( <i>dd</i> , $J = 15.3, 9.0$ )	C(12), C(13), C(14), C(16)
$\text{H}_\beta\text{-C}(15)$		3.01 ( <i>dd</i> , $J = 15.3, 7.0$ )	
H–C(16)	86.4 ( <i>d</i> )	5.12–5.16 ( <i>m</i> )	C(15), C(17)
$\text{H}_\alpha\text{-C}(17)$	64.8 ( <i>t</i> )	3.89 ( <i>dd</i> , $J = 15.0, 8.7$ )	C(15), C(16)
$\text{H}_\beta\text{-C}(17)$		3.80 ( <i>dd</i> , $J = 15.0, 7.2$ )	
Me(18)	29.6 ( <i>q</i> )	1.40 ( <i>s</i> )	C(3), C(4), C(5), C(19)
Me(19)	20.1 ( <i>q</i> )	1.44 ( <i>s</i> )	C(3), C(4), C(5), C(18)
Me(20)	26.6 ( <i>q</i> )	1.46 ( <i>s</i> )	C(1), C(5), C(9), C(10)
11-OH		5.41 (br. <i>s</i> )	
14-OH		13.34 (br. <i>s</i> )	

<sup>a</sup>) Assignments were accomplished by a combination of 1D- and 2D- ( $^1\text{H}$ ,  $^1\text{H}$ -COSY, HSQC, and HMBC) NMR experiments.

**Biological Studies.** Compounds **1** and **2** were evaluated for their cytotoxic activities against the HL-60 and A-549 cell lines *in vitro* by means of the MTT (= 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide) assay [10]. Both compounds were found to be significantly active to inhibit the proliferation of HL-60 and A-549 cell lines with  $IC_{50}$  values less than 15  $\mu\text{M}$ , as compared to cisplatin used as positive control (Table 3).

Table 3. Cytotoxicities ( $IC_{50}$  [ $\mu\text{M}$ ]) of Compounds **1** and **2** Isolated from *Clerodendrum kaichianum* P. S. HSU

Compound	HL-60	A-549
<b>1</b>	5.95 $\pm$ 0.57	9.37 $\pm$ 0.24
<b>2</b>	15.91 $\pm$ 0.95	10.35 $\pm$ 0.51
Cisplatin	4.87 $\pm$ 0.89	8.63 $\pm$ 1.19

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### Experimental Part

**General.** Column chromatography (CC): silica gel ( $\text{SiO}_2$ , 200–300 mesh; *Qingdao Marine Chemical Factory*, Qingdao, China) and *Sephadex LH-20* (*Amersham Pharmacia Biotech*, UK). Prep. HPLC: *JAI-9103* (*Japan Analytical Industry*). TLC: *Merck* silica-gel plates; visualization under UV light and by spraying with 10% aq.  $\text{H}_2\text{SO}_4$ , followed by heating. M.p.: *X-4* apparatus; uncorrected. Optical rotations: *Perkin-Elmer 241* automatic polarimeter. UV Spectra: *Shimadzu UV-2550* spectrophotometer, in MeOH;  $\lambda_{\text{max}}$  (log  $\epsilon$ ) in nm. IR Spectra: *Nicolet 380 FT-IR* spectrophotometer (*Thermo Nicolet*); KBr pellets; in  $\text{cm}^{-1}$ .  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR: *Bruker AVANCE III 500* spectrometer, at 500 ( $^1\text{H}$ ), and 125 ( $^{13}\text{C}$ ) MHz; in  $\text{CDCl}_3$ ;  $\delta$  in ppm,  $J$  in Hz. HR-ESI-MS: *Agilent 6210* TOF-MS mass spectrometer.

**Plant Material.** The stems of *C. kaichianum* were collected on the mountains of Lin'an County, Zhejiang Province, P. R. China, in September of 2009, and identified by Dr. *Bin Wu* (College of Pharmaceutical Sciences, Zhejiang University, Hangzhou, China). A voucher specimen (No. 20090913) was deposited with the laboratory of Zhejiang Gongshang University, Hangzhou, P. R. China.

**Extraction and Isolation.** The air-dried powder of the stems (11.6 kg) of *C. kaichianum* was extracted with 75% aq. EtOH three times at 75° for 4 h each. The EtOH extracts were combined and evaporated to dryness to afford a gummy residue (325 g), which was suspended in  $\text{H}_2\text{O}$  (2 l) and extracted at r.t. with petroleum ether (PE; 3  $\times$  2 l, 65 g), AcOEt (3  $\times$  2 l, 102 g), and BuOH (3  $\times$  2 l, 88 g), successively.

Part of AcOEt extract (90 g) was subjected to CC ( $\text{SiO}_2$ ; AcOEt/PE gradient) to afford ten fractions, *Frs. 1–10*, on the basis of TLC analysis. *Fr. 2* (8 g) was further subjected CC ( $\text{SiO}_2$ ; AcOEt/PE 5–15%) and separated into five fraction pools, *Frs. 2A, 2B, 2C, 2D, 2E*. *Fr. 2D* was further purified on *Sephadex LH-20* and later prep. HPLC ( $\text{H}_2\text{O}/\text{MeOH}$  60 : 40) to afford compounds **3** (12 mg), **4** (40 mg), **5** (17 mg), and **6** (9 mg).

The major fraction, *Fr. 4* (9 g), was further subjected CC ( $\text{SiO}_2$ ; PE/acetone from 5 : 1 to 1 : 1) to give six fraction pools, *Frs. 4A, 4B, 4C, 4D, 4E, 4F*. *Fr. 4E* was subjected to *Sephadex LH-20*, and later prep. HPLC to give compounds **1** (8 mg) and **2** (11 mg).

**17-Hydroxyteuvincenone G** (= (16R)-12,16-Epoxy-11,14,17-trihydroxy-17(15  $\rightarrow$  16)-abeo-abieta-8,11,13-triene-2,7-dione; **1**). Yellowish needles. M.p. 250–252°.  $[\alpha]_{\text{D}}^{25} = +150.1$  ( $c = 0.02$ , MeOH); UV(MeOH): 207, 225, 280, 360. IR: 3400, 2992, 1717, 1650, 1615, 1699, 1581.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR ( $\text{CDCl}_3$ ): see Table 1. HR-ESI-MS: 359.1487 ( $[M - \text{H}]^-$ ,  $\text{C}_{20}\text{H}_{23}\text{O}_6^-$ ; calc. 359.1494).

**17-Hydroxyteuvincen-5(6)-enone G** (= (16R)-12,16-Epoxy-11,14,17-trihydroxy-17(15  $\rightarrow$  16)-abeo-abieta-5,8,11,13-tetraene-2,7-dione; **2**). Yellowish needles. M.p. 305–307°.  $[\alpha]_{\text{D}}^{25} = +47.2$  ( $c = 0.07$ ,

MeOH). UV(MeOH): 203, 260, 299, 365. IR: 3396, 2980, 1716, 1650, 1622, 1693, 1679. <sup>1</sup>H- and <sup>13</sup>C-NMR (CDCl<sub>3</sub>): see Table 2. HR-ESI-MS: 357.1330 ([M – H]<sup>–</sup>, C<sub>20</sub>H<sub>21</sub>O<sub>6</sub><sup>–</sup>; calc. 357.1336).

The percentage of growth inhibition was determined using a MTT assay to measure viable cells with minor modification. A total of 5000–10000 exponential phase cells per well was seeded onto a 96-well plate for 24 h, treated with compounds **1** and **2** at 2.5, 5, 10, 20, 40 μM concentrations for 72 h using cisplatin as a positive control. Briefly, MTT (= 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide; 100 μl) working soln. (1 mg/ml) was added into each well of 96-well plate and incubated at 37° for 4 h, and then the medium was removed. The converted dye formazan was solubilized with 150 μl acidic i-PrOH, and each concentration was tested in triplicate. The absorbance was then measured at a wavelength of 570 nm using a microplate reader. The dose resulting in 50% inhibition of cell growth, IC<sub>50</sub>, was calculated by NDST software.

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